

Gram proportion determination in complex ecosystems using flow cytometry

Aurore DUQUENOY^{1,2}, Samuel BELLAIS², Vincent THOMAS²
¹MaaT Pharma – Lyon – France, ²Bioaster – Paris - France

Contact: aduquenoy@maat-pharma.com

INTRODUCTION

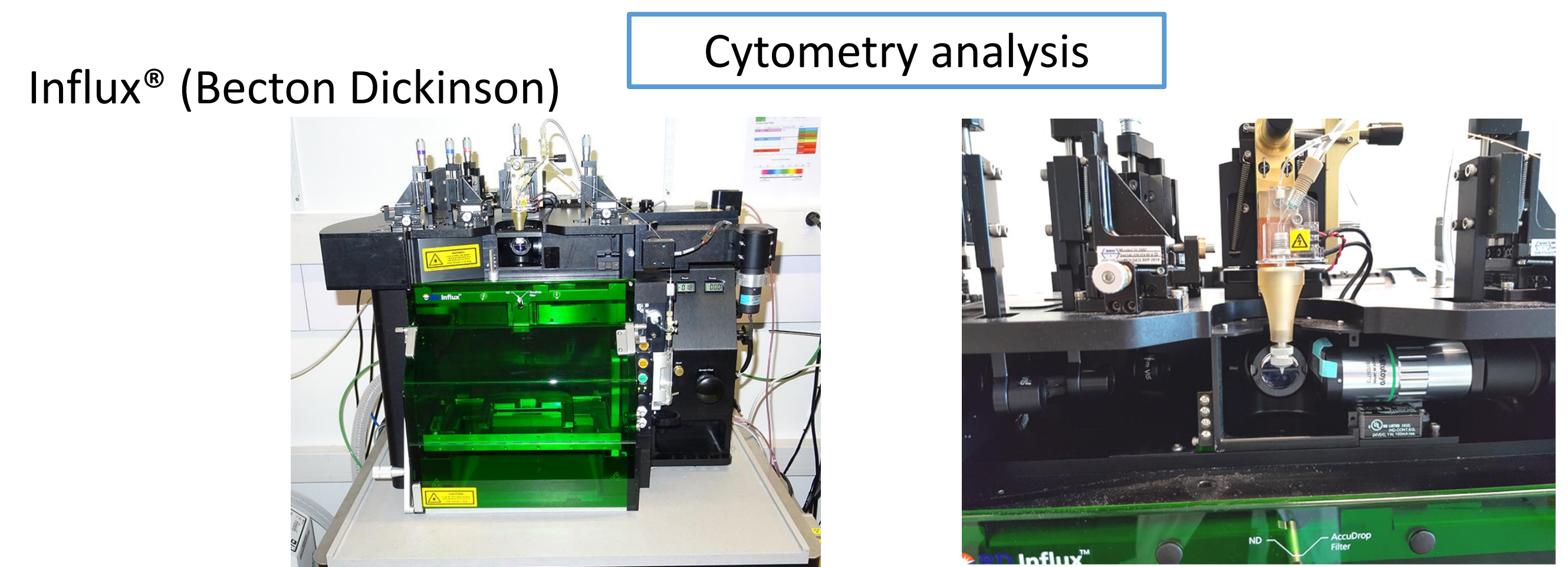
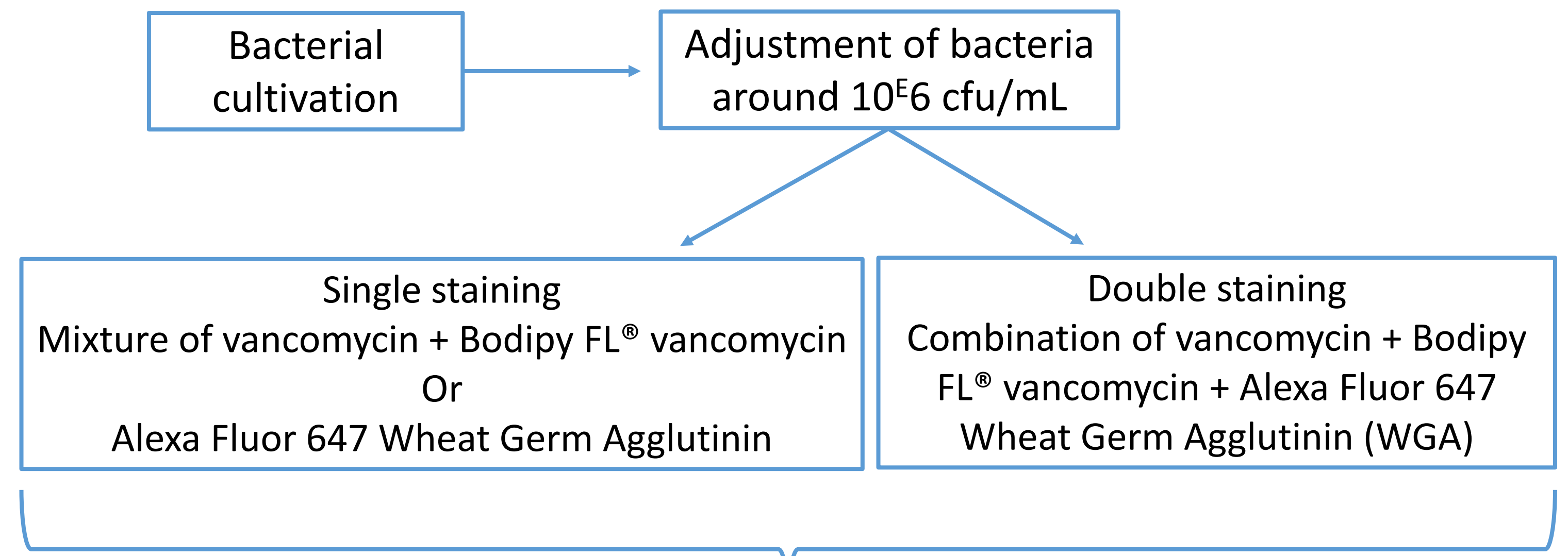
A first glimpse of the evolution of a complex ecosystem during *in vitro* fermentation can be assessed using flow cytometry methods. General bacterial profiles, overall viability as well as Gram status can be evaluated.

Here, we adapted a Gram-staining method using flow cytometry for *in vitro* ecosystems characterization. The method was developed and validated using pure cultures of aerobic bacterial species and then applied to anaerobic species and complex ecosystems.

OBJECTIVES

The objective was to develop a Gram staining method adapted for the analysis of complex ecosystems. We aimed at evaluating the most appropriate staining conditions to discriminate Gram-positive from Gram-negative bacteria. Aerobic as well as strictly anaerobic strains were tested.

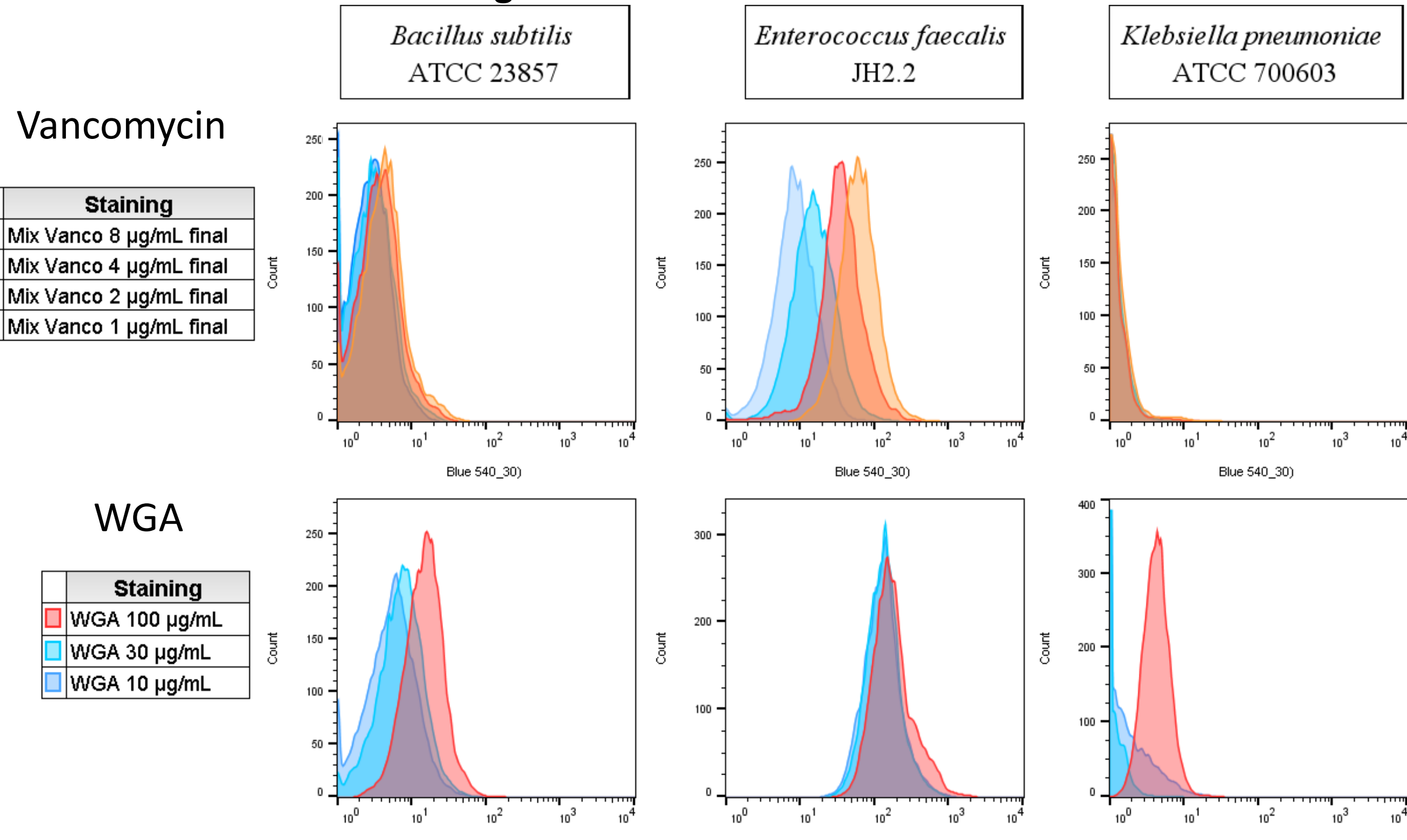
METHODS



RESULTS

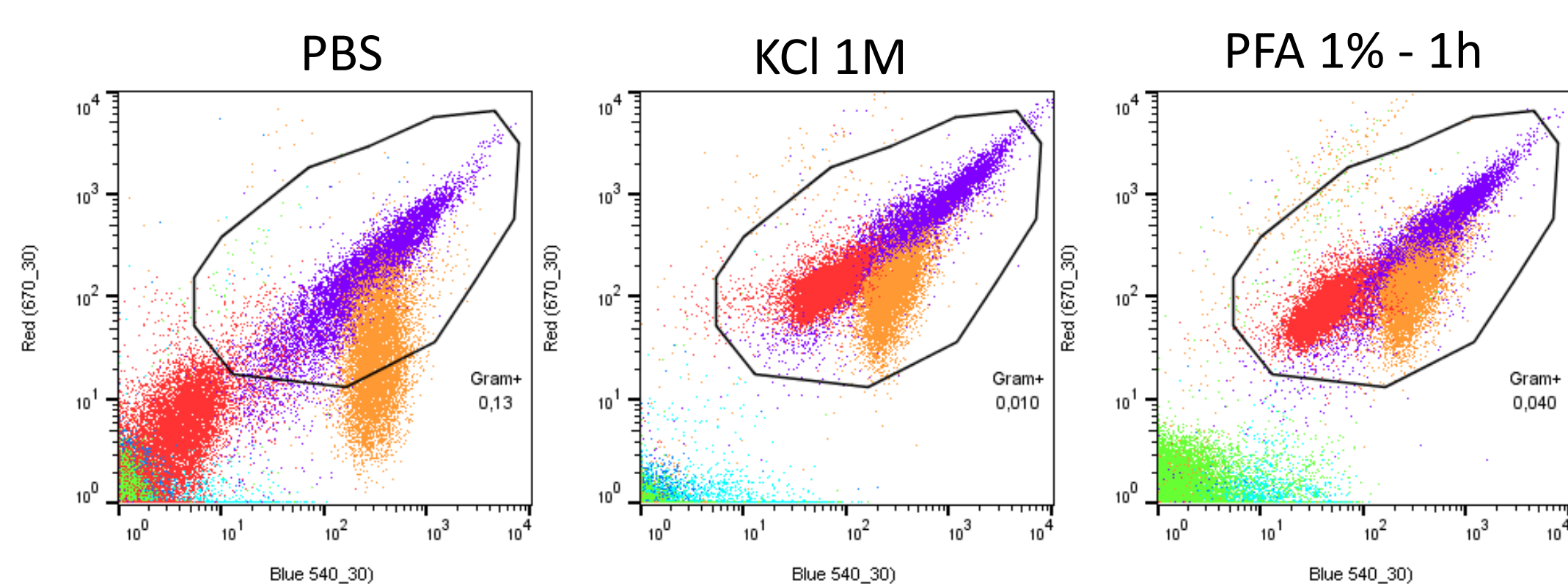
Optimization of Gram-positive and Gram-negative bacteria differentiation

Stains concentrations investigation



- Vancomycin**
- Gram-positive bacteria: higher concentrations **increased** the staining
 - Gram-negative bacteria: **no staining**
- WGA**
- Better staining** of Gram-positive bacteria with increased concentrations
 - Higher concentrations: unwanted Gram-negative bacteria staining

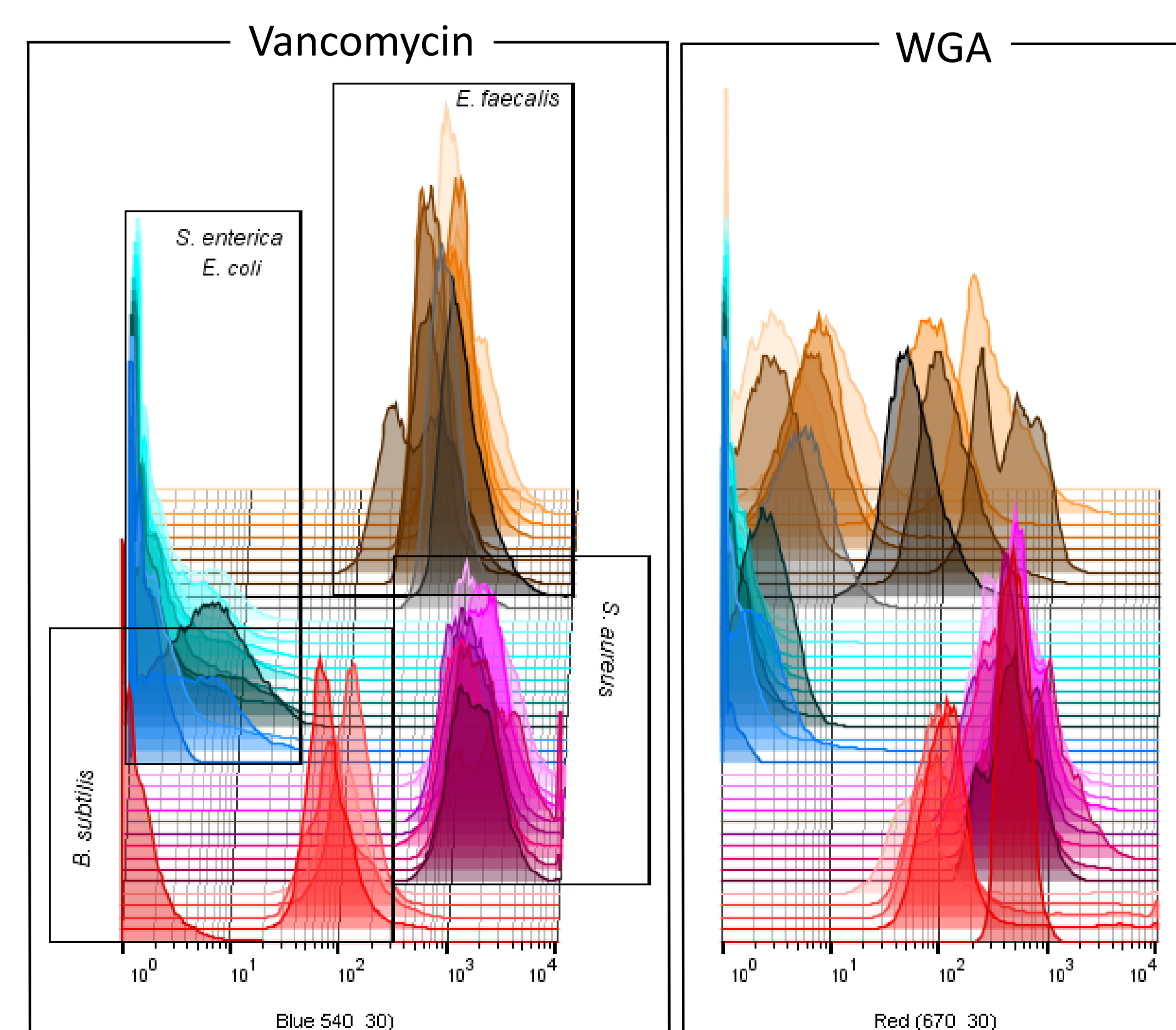
Stains combination and fixation condition



green: *K. pneumoniae* ATCC 700603, light blue: *Salmonella enterica* ATCC 13311, dark blue: *Escherichia coli* ATCC 35218, orange: *E. faecalis* ATCC 29212, purple: *Lactococcus lactis* ATCC 11454, red: *B. subtilis* ATCC 23857

KCl treatment improves Gram-positive bacteria staining leading to a better discrimination from Gram-negative bacteria especially for *B. subtilis*. In addition, the PFA fixation method after KCl treatment does not impact the staining.

Screening of different strains of *E. faecalis*, *B. subtilis*, *S. aureus*, *L. lactis*, *E. coli* and *S. enterica*

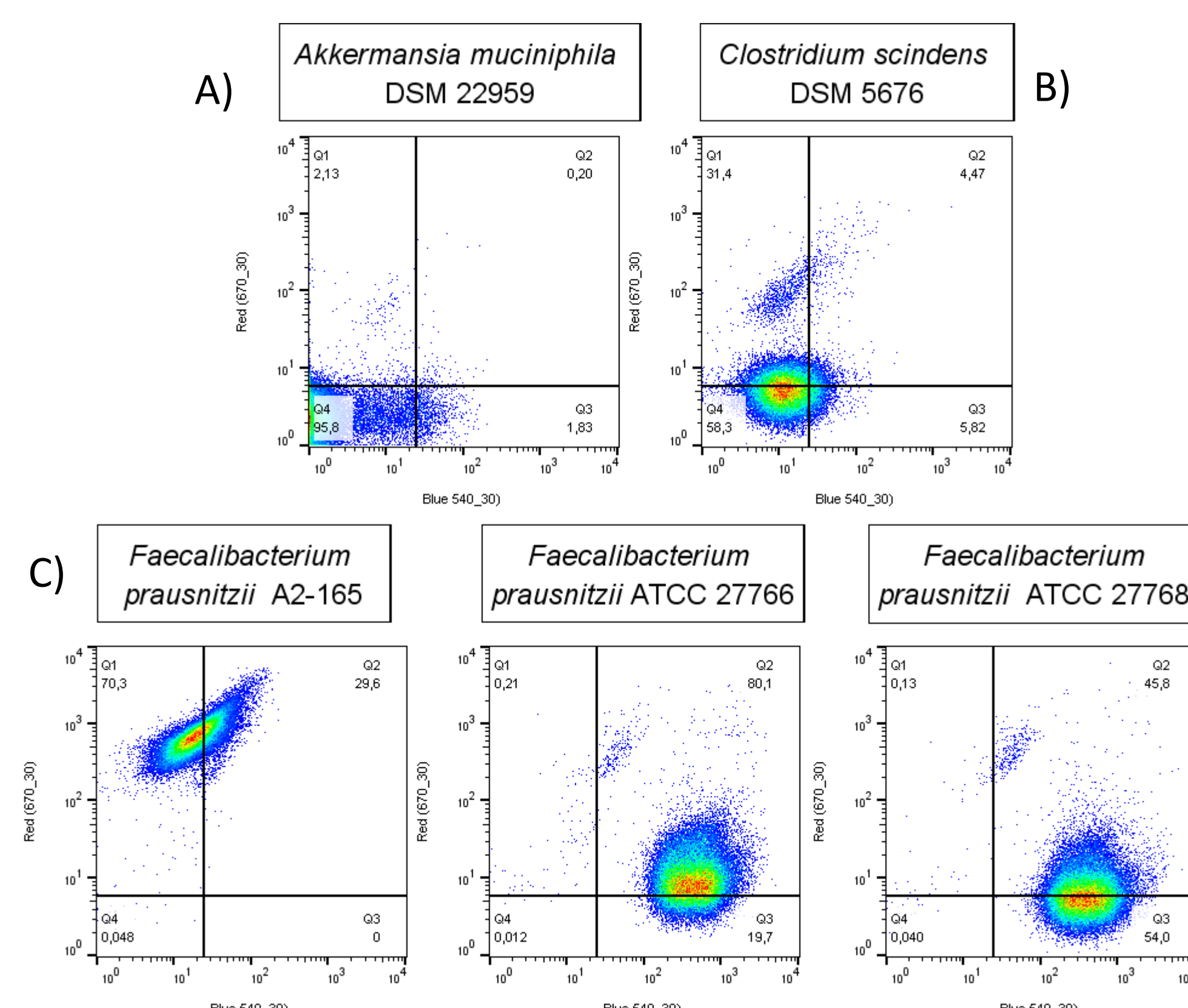


- Vancomycin**
- Gram -: **not stained**
 - Gram +: almost always stained (one strain of *B. subtilis* not stained)

- WGA**
- Gram -: **not stained**
 - Gram+: heterogeneous staining depending on the tested strains

➤ The **combination of both stains** represents an interesting option for Gram-positive bacteria that might not be stained by one of the selected stains

Screening of different anaerobic strains



- A) *A. muciniphila* is stained as a Gram-negative bacteria as expected
- B) *C. scindens* appears to be stained as a Gram-positive bacterium
- C) The three *F. prausnitzii* strains are stained as Gram-positive bacteria. A significant difference is observed between the A2-165 strain and the two ATCC strains that might be due to differences on cell walls composition.

CONCLUSIONS

- Differentiation of Gram-positive bacteria and Gram-negative bacteria based on their cell wall structures
- Under the conditions and on the strains tested, gram differentiation was possible whatever bacterial growth phases (data not shown)
- Observation of staining variability for different strains belonging to a same species (likely due to differences in proteins expressed at the surface of bacteria)
- Gram-positive bacteria could always be differentiated from Gram-negative bacteria
- Gram-staining method that may be used in the future to predict the presence of specific bacterial groups in *in vitro* cultures as well as in complex ecosystems